WILEY

DOI: 10.1111/acel.14352

SHORT COMMUNICATION

Aging Cell

Emerging signs of Alzheimer-like tau hyperphosphorylation and neuroinflammation in the brain post recovery from COVID-19

¹Neuroscience Research Institute and Department of Neurobiology, School of Basic Medical Sciences, Peking University, Beijing, China

²Key Laboratory for Neuroscience, Ministry of Education/National Health Commission, Peking University, Beijing, China

 3 Key Laboratory of Human Brain Bank for Functions and Diseases of Department of Education of Guizhou Province, Guizhou Medical University, Guiyang, China

4 Key Laboratory of Endemic and Ethnic Diseases, Ministry of Education, Guizhou Medical University, Guiyang, China

5 School of Forensic Medicine, Guizhou Medical University, Guiyang, China

6 Beijing Life Science Academy, Beijing, China

Correspondence

You Wan and Jie Zheng, Neuroscience Research Institute and Department of Neurobiology, School of Basic Medical Sciences, Peking University, Beijing 100083, China.

Email: ywan@hsc.pku.edu.cn and zhengjiie@hsc.pku.edu.cn

Funding information

Peking University Talent Startup Fund, Grant/Award Number: BMU2022YJ003; Fundamental Research Funds for the Central Universities; Scientific Project of Beijing Life Science Academy, Grant/ Award Number: 2023000CA0060, 2023000CB0010 and 2024300CC0020; Guizhou Provincial Science and Technology Program Project Grant, Grant/Award Number: Qiankehe Platform Talents-CXTD[2023]003

Abstract

Coronavirus disease 2019 (COVID-19) has been suggested to increase the risk of memory decline and Alzheimer's disease (AD), the main cause of dementia in the elderly. However, direct evidence about whether COVID-19 induces AD-like neuropathological changes in the brain, especially post recovery from acute infection, is still lacking. Here, using postmortem human brain samples, we found abnormal accumulation of hyperphosphorylated tau protein in the hippocampus and medial entorhinal cortex within 4–13 months post clinically recovery from acute COVID-19, together with prolonged activation of glia cells and increases in inflammatory factors, even though no SARS-COV-2 invasion was detected in these regions. By contrast, COVID-19 did not change beta-amyloid deposition and hippocampal neuron number, and had limited effects on AD-related pathological phenotypes in olfactory circuits including olfactory bulb, anterior olfactory nucleus, olfactory tubercle, piriform cortex and lateral entorhinal cortex. These results provide neuropathological evidences linking COVID-19 with prognostic increase of risk for AD.

KEYWORDS

Alzheimer's disease, COVID-19, glia cell, inflammation, tau

Abbreviations: AD, Alzheimer's disease; AON, anterior olfactory nucleus; Aβ, amyloid-beta; COVID-19, Coronavirus disease 2019; ddPCR, droplet digital PCR; GFAP, glial Fibrillary acidic protein; HMGB1, high mobility group box 1; IL-10, interleukin-10; IL-18, interleukin-18; IL-1β, interleukin-1β; IL-6, interleukin-6; LEC, lateral entorhinal cortex; MEC, medial entorhinal cortex; NAAT, nucleic acid amplification test; OB, olfactory bulb; OT, olfactory tubercle; P1~P4, pattern 1~4; PAI1, plasminogen activator inhibitor-1; Pir, piriform cortex; pTau, phosphorylated tau; TNF-α, tumor necrosis factor alpha.

Xuetao Qi, Shulu Yuan and Jiuyang Ding contributed equally to this work.

This is an open access article under the terms of the Creative Commons [Attribution](http://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2024 The Author(s). *Aging Cell* published by Anatomical Society and John Wiley & Sons Ltd.

2 of 6 | WILEY- Aging Cell (a) **Contract of the CONSTANT** QUETAL.

Cognitive decline including memory problems, mental cloudiness and difficulty in concentrating (collectively known as "brain fog") has been widely recognized as parts of neuropathological sequelae of coronavirus disease 2019 (COVID-19) in recent years (Thaweethai et al., [2023](#page-5-0)), and captures increasing attention on whether and how the history of Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection would increase the risk of Alzheimer's disease (AD) in later life, especially for the elderly (Bonhenry et al., [2024;](#page-4-0) Olivera et al., [2023](#page-5-1)). However, direct evidence on the appearance of AD-related neuropathological features, like abnormal accumulation of hyperphosphorylated tau protein and amyloid-beta (Aβ) as well as glia dysfunction and neuroinflammation in the brain (Long & Holtzman, [2019](#page-5-2)), in human brain post recovery from acute COVID-19 is still in lack.

Here, we collected postmortem human brain samples from individuals died with SARS-COV-2 non-infected (group "N", *n*= 6), during acute COVID-19 (group "C", $n=6$), and within 4-13 months post-clinically recovered from COVID-19 (group "R", *n*= 7). Only individuals who had no cognitive impairment reported before SARS-COV-2 infection were included, and the infection and recovery of SARS-COV-2 was determined or ruled out by nucleic acid amplification test (NAAT). Individual information was summarized in Table [S1](#page-5-3).

We first examined the level of phosphorylated tau (pTau) in the hippocampus and medial entorhinal cortex (MEC), since these regions play pivotal roles in the development of AD, and have been well-recognized to be vulnerable for pTau accumulation and neurodegeneration (Braak et al., [2006](#page-4-1)). Tau species ranging from 35 to 100 kD and phosphorylated at multiple AD-related epitopes were measured. Interestingly, we found that both pTau and total tau showed nonsignificant change in patients died during acute COVID-19 compared with individuals without SARS-COV-2 infection, but pTau was unexpectedly upregulated in the group clinically recovered from COVID-19, especially at AD-related epitopes like Thr181, Thr217 and AT8 (Figure [1a–c,](#page-2-0) Figure [S1\)](#page-5-3).

Meanwhile, we also evaluated the distribution pattern of pTau in the hippocampus and MEC, which were classified here into patterns 1–4 (P1–P4) based on the immunohistochemical staining intensity (supplementary methods—Data [S1](#page-5-3)). The post-acute COVID-19 group generally exhibited higher degree of pTau aggregation compared with SARS-COV-2 non-infected controls at Thr181 and Thr217 epitopes (Figure [1d,e](#page-2-0)).

We next examined two other AD-related pathologies including Aβ deposition and hippocampal neuron loss. By contrast to tau, no statistical change in the density and size of Aβ plaques was measured (Figure 1f-h), and no hippocampal neuron loss was found both during and post-acute COVID-19 (Figure [1i,j](#page-2-0)).

Besides, taken into consideration that COVID-19 and earlystage AD shared symptom of smell loss (Doty, [2022](#page-4-2); Tsukahara et al., [2023](#page-5-4)), a newly-identified biomarker of cognitive impairment (Murphy, [2019](#page-5-5)), we wonder whether acute or post-acute COVID-19 affects pTau and Aβ in the olfactory circuit consisting of olfactory bulb (OB), anterior olfactory nucleus (AON), olfactory tubercle (OT),

piriform cortex (Pir) and lateral entorhinal cortex (LEC). By contrast to the hippocampus and MEC, no significant change in pTau and Aβ plaque was measured in all olfactory areas (Figure [S2](#page-5-3)).

Subsequently, we sought to explore whether the elevation of pTau was associated with potential invasion of SARS-COV-2 into the brain, which was found to be capable of directly inducing tau hyperphosphorylation in cultured neuroblastoma cells and 3D human brain organoids (Di Primio et al., [2023](#page-4-3); Ramani et al., [2020](#page-5-6)). However, we did not observe the existence of SARS-COV-2 in the brain utilizing immunohistochemical staining of nucleocapsid (N) and spike glycoprotein (S) protein (Figure [S3\)](#page-5-3). Consistently, we did not detect SARS-COV-2 ORFab1 and N protein mRNA by both real-time quantitative reverse transcription PCR (RT-qPCR) and droplet digital PCR (ddPCR) (Figures [S4](#page-5-3) and [S5](#page-5-3)).

Given an important contribution of abnormal glia activation and neuroinflammation to the elevation of pTau in the development of AD (Leng & Edison, [2021](#page-5-7)), we measured whether COVID-19 induced glia dysfunction in the hippocampus and MEC, and found increased soma volume of and less ramified processes of microglia during and post-acute COVID-19, two morphological signs indicating the activation microglia (Leng & Edison, [2021](#page-5-7); Woodburn et al., [2021](#page-5-8)), though microglia did not change in number (Figure [2a–e](#page-3-0)). Similarly, the number of GFAP-labeled astrocytes did not change, but both the soma volume and complexity of processes of astrocytes increased post recovery from COVID-19 compared with acute COVID-19 (Figure [2f–j](#page-3-0)), indicating that astrocytes were also activated during and post COVID-19. Consistently, the activation of glia cells was also indicated by the upregulation in protein levels of Iba1, CD68, GFAP and S100β in groups of acute and post-acute COVID-19 (Figure [S6](#page-5-3)).

Moreover, the expression levels of inflammatory factors including tumor necrosis factor alpha (TNF-α), interleukin-1β (IL-1β) and interleukin-6 (IL-6) were all upregulated in acute and post-acute COVID-19, though other cytokines like interleukin-10 (IL-10) and interleukin-18 (IL-18), as well as neuroinflammation-related proteins like high mobility group box 1 (HMGB1) and plasminogen activator inhibitor-1 (PAI1) remained nearly unchanged (Figure [2k](#page-3-0)).

In line with our results, the risk of AD has been recently found to increase post-acute COVID-19 (Bonhenry et al., [2024](#page-4-0)). A follow-up study found persist elevation of AD-related plasma biomarkers including total-tau, pTau181, inflammatory cytokines, NfL, neurogranin, etc. at 1 to 3 months after initial SARS-CoV-2 infection (Sun et al., [2021](#page-5-9)). Meanwhile, following with structural changes in brain during acute COVID-19 (Douaud et al., [2022](#page-4-4)), persistent white matter changes in the brain of people recovered from COVID-19 were also reported at 1-year follow-up (Huang et al., [2022](#page-4-5)). These findings suggest the existence of long-term brain neuropathology after COVID-19, which might be at least partly responsible for the postacute sequelae of cognition and memory impairment (Hampshire et al., [2024](#page-4-6)).

We sought to investigate in the present study preliminarily the potential cause of pTau elevation afterCOVID-19. In discrepancy with several previous studies (Emmi et al., [2023](#page-4-7); Gagliardi et al., [2021;](#page-4-8) Stein et al., [2022](#page-5-10)), we did not detect SARS-COV-2 particles in all

Licens

FIGURE 1 Upregulation of pTau in the hippocampus and MEC post COVID-19. (a–c) pTau at specific epitopes like Thr181, Thr217 and AT8 were upregulated in the hippocampus and MEC post-acute COVID-19. Grouping: N, non-infected; C, acute COVID-19; R, recovery from COVID-19. *N*= 6–7 in each group, one-way ANOVA followed by Tukey's multiple comparisons tests. Data were normalized by the mean value of group N for each pTau epitope. Extended blots were shown in Figure [S1](#page-5-3). (d, e) Measurement of pTau distribution pattern in the hippocampus and MEC indicated increase of pTau aggregation post-acute COVID-19. Representative images showed pattern 1-4 (P1-P4) of pTau accumulation for each epitope and in each group (d). Numbers in pie charts indicated the counts of patients for each pattern (e). *N*= 6–7 in each group. (f–h) The overall density and averaged size of 6E10-stained Aβ plaques did not change in acute and post-acute COVID-19. *N*= 6–7 in each group, one-way ANOVA followed by Tukey's multiple comparisons tests. (i, j) Neuronal density in each hippocampal subregion did not change in acute and post-acute COVID-19. *N*= 6–7 individuals in each group, one-way ANOVA followed by Tukey's multiple comparisons tests.

FIGURE 2 Prolonged upregulation of glia activation and inflammatory factors expression in the hippocampus post-acute COVID-19. (a, b) Representative immunofluorescent images (a) and 3D-reconstruacted morphology (b) of Iba1-stained microglia. (c–e) The number of microglia did not change (c), but soma volume significantly increased (d) and branches complexity decreased (e) in acute and post-acute COVID-19. *N*= 6–7 patients or 174–382 cells in each group, ****p*< 0.001, one-way (c, d) and repeated measures (e) ANOVA followed by Tukey's multiple comparisons tests. (f, g) Representative immunohistochemical images (f) and Sholl analysis diagram (g) of GFAP-stained astrocytes. (h–j) The number of astrocytes did not change (h), but both soma volume (i) and processes complexity (j) significantly increased in acute and post-acute COVID-19. *N*= 6–7 patients or 108–194 cells in each group, ****p*< 0.001, one-way ANOVA followed by Tukey's multiple comparisons tests. (k) mRNA levels of inflammatory factors including TNF-α, IL-1β and IL-6 were upregulated in acute and post-acute COVID-19. *N*= 6–7 patients in each group, one-way ANOVA followed by Tukey's multiple comparisons tests.

brain regions measured in the present study. Instead, we revealed in individuals post-acute COVID-19 prolonged glia activation and neuroinflammation, two other important drivers of tau hyperphosphorylation in the development of AD (Chen & Yu, [2023;](#page-4-9) van der Kant et al., [2020](#page-5-11)). In fact, increased glia activation and inflammatory factors have been widely-recognized both in the brain and plasma of COVID-19 patients (Chen et al., [2020](#page-4-10); Huang et al., [2020](#page-4-11); Matschke

et al., [2020](#page-5-12); Zhang et al., [2023](#page-5-13)). The central neuroinflammatory infiltration might be a secondary consequence of the peripheral cytokine storm during acute SARS-CoV-2 infection due to the damage of brain–blood barrier (Amruta et al., [2022](#page-4-12); Schwabenland et al., [2021](#page-5-14)). Excessive activation of glia cells and inflammation in the brain can exacerbate tau phosphorylation and aggregation by dysregulating tau kinases and phosphatases (Ising et al., [2019](#page-4-13)).

Nevertheless, our pilot study only included a small number of postmortem samples, it deserves further investigation to collect data from larger cohorts. Besides, it also remains to be elucidated whether and how long these AD-associated pathological phenotypes persist following years to decades after recovery from acute COVID-19, including the increased pTau level and neuroinflammation found here. The prognostic risk of AD might be better indicated through long-term measurement of plasma or cerebrospinal fluid biomarkers (Ossenkoppele et al., [2022](#page-5-15)), brain imaging of early-AD fea-tures (Jagust, [2018\)](#page-4-14), and cognition-testing scales (Woo et al., [2020](#page-5-16)).

In conclusion, we reported here emerging signs of AD-like elevation of pTau and neuroinflammation during and post-acute COVID-19 in the hippocampus and MEC of postmortem human brain, which provided neuropathological evidences about increased risk of AD in long COVID-19.

AUTHOR CONTRIBUTIONS

Conceptualization: X. Q., J. D., J. Z; *Methodology*: X. Q., S. Y., J. D., Y. Y., S. F; *Investigation*: X. Q., S. Y., J. D., W. S., Y. S., Y. X., Z. L; *Data analysis*: X. Q., S. Y., J. Z; *Manuscript writing*: X. Q., J. Z; *Manuscript revision*: X. Q., B. S., X. Q., B. X., F. L., M. Y., J. M., Y. W., J.Z; *Funding acquisition*: J. M., Y. W., J. Z; *Resources*: J. D., Y. W., J. Z; *Supervision*: B. S., X. Q., B. X., Y. W., J. Z.

ACKNOWLEDGEMENTS

This work was supported in part by the Peking University Talent Startup Fund (BMU2022YJ003) supported by the "Fundamental Research Funds for the Central Universities," Scientific Project of Beijing Life Science Academy (2024300CC0020, 2023000CA0060, 2023000CB0010), and Guizhou Provincial Science and Technology ProgramProjectGrant(QiankehePlatformTalents-CXTD[2023]003).

CONFLICT OF INTEREST STATEMENT

All authors declare no conflicts interest.

DATA AVAILABILITY STATEMENT

All data were available by reasonable requirements to Dr. Jie Zheng (zhengjiie@hsc.pku.edu.cn).

MATERIALS AND METHODS

Please see supplementary materials—Data [S1](#page-5-3).

ORCID

Xuetao Qi <https://orcid.org/0000-0003-1449-7268> *Xiaolan Qi* <https://orcid.org/0000-0002-9108-6772> *Jie Zhen[g](https://orcid.org/0000-0002-0637-9918)* <https://orcid.org/0000-0002-0637-9918>

REFERENCES

Amruta, N., Ismael, S., Leist, S. R., Gressett, T. E., Srivastava, A., Dinnon, K. R., Engler-Chiurazzi, E. B., Maness, N. J., Qin, X., Kolls, J. K., Baric, R. S., & Bix, G. (2022). Mouse adapted SARS-CoV-2 (MA10) viral infection induces neuroinflammation in standard laboratory mice. *Viruses*, *15*, 15.

Aging Cell \bullet **Cell** \bullet

- Bonhenry, D., Charnley, M., Goncalves, J., Hammarstrom, P., Heneka, M. T., Itzhaki, R., Lambert, J. C., Mannan, M., Baig, A. M., Middeldorp, J., Nyström, S., Reynolds, N. P., Stefanatou, M., & Berryman, J. T. (2024). SARS-CoV-2 infection as a cause of neurodegeneration. *Lancet Neurology*, *23*, 562–563.
- Braak, H., Alafuzoff, I., Arzberger, T., Kretzschmar, H., & Del, T. K. (2006). Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathologica*, *112*, 389–404.
- Chen, G., Wu, D., Guo, W., Cao, Y., Huang, D., Wang, H., Wang, T., Zhang, X., Chen, H., Yu, H., Zhang, X., Zhang, M., Wu, S., Song, J., Chen, T., Han, M., Li, S., Luo, X., Zhao, J., & Ning, Q. (2020). Clinical and immunological features of severe and moderate coronavirus disease 2019. *The Journal of Clinical Investigation*, *130*, 2620–2629.
- Chen, Y., & Yu, Y. (2023). Tau and neuroinflammation in Alzheimer's disease: Interplay mechanisms and clinical translation. *Journal of Neuroinflammation*, *20*, 165.
- Di Primio, C., Quaranta, P., Mignanelli, M., Siano, G., Bimbati, M., Scarlatti, A., Piazza, C. R., Spezia, P. G., Perrera, P., Basolo, F., Poma, A. M., Costa, M., Pistello, M., & Cattaneo, A. (2023). Severe acute respiratory syndrome coronavirus 2 infection leads to tau pathological signature in neurons. *PNAS Nexus*, *2*, pgad282.
- Doty, R. L. (2022). Olfactory dysfunction in COVID-19: Pathology and long-term implications for brain health. *Trends in Molecular Medicine*, *28*, 781–794.
- Douaud, G., Lee, S., Alfaro-Almagro, F., Arthofer, C., Wang, C., McCarthy, P., Lange, F., Andersson, J., Griffanti, L., Duff, E., Jbabdi, S., Taschler, B., Keating, P., Winkler, A. M., Collins, R., Matthews, P. M., Allen, N., Miller, K. L., Nichols, T. E., & Smith, S. M. (2022). SARS-CoV-2 is associated with changes in brain structure in UK biobank. *Nature*, *604*, 697–707.
- Emmi, A., Rizzo, S., Barzon, L., Sandre, M., Carturan, E., Sinigaglia, A., Riccetti, S., Della, B. M., Boscolo-Berto, R., Cocco, P., Macchi, V., Antonini, A., De Gaspari, M., Basso, C., De Caro, R., & Porzionato, A. (2023). Detection of SARS-CoV-2 viral proteins and genomic sequences in human brainstem nuclei. *NPJ Parkinsons Dis*, *9*, 25.
- Gagliardi, S., Poloni, E. T., Pandini, C., Garofalo, M., Dragoni, F., Medici, V., Davin, A., Visona, S. D., Moretti, M., Sproviero, D., Pansarasa, O., Guaita, A., Ceroni, M., Tronconi, L., & Cereda, C. (2021). Detection of SARS-CoV-2 genome and whole transcriptome sequencing in frontal cortex of COVID-19 patients. *Brain, Behavior, and Immunity*, *97*, 13–21.
- Hampshire, A., Azor, A., Atchison, C., Trender, W., Hellyer, P. J., Giunchiglia, V., Husain, M., Cooke, G. S., Cooper, E., Lound, A., Donnelly, C. A., Chadeau-Hyam, M., Ward, H., & Elliott, P. (2024). Cognition and memory after Covid-19 in a large community sample. *The New England Journal of Medicine*, *390*, 806–818.
- Huang, C., Wang, Y., Li, X., Ren, L., Zhao, J., Hu, Y., Zhang, L., Fan, G., Xu, J., Gu, X., Cheng, Z., Yu, T., Xia, J., Wei, Y., Wu, W., Xie, X., Yin, W., Li, H., Liu, M., … Cao, B. (2020). Clinical features of patients infected with 2019 novel coronavirus in Wuhan China. *Lancet*, *395*, 497–506.
- Huang, S., Zhou, Z., Yang, D., Zhao, W., Zeng, M., Xie, X., Du, Y., Jiang, Y., Zhou, X., Yang, W., Guo, H., Sun, H., Liu, P., Liu, J., Luo, H., & Liu, J. (2022). Persistent white matter changes in recovered COVID-19 patients at the 1-year follow-up. *Brain*, *145*, 1830–1838.
- Ising, C., Venegas, C., Zhang, S., Scheiblich, H., Schmidt, S. V., Vieira-Saecker, A., Schwartz, S., Albasset, S., McManus, R. M., Tejera, D., Griep, A., Santarelli, F., Brosseron, F., Opitz, S., Stunden, J., Merten, M., Kayed, R., Golenbock, D. T., Blum, D., … Heneka, M. T. (2019). NLRP3 inflammasome activation drives tau pathology. *Nature*, *575*, 669–673.
- Jagust, W. (2018). Imaging the evolution and pathophysiology of Alzheimer disease. *Nature Reviews. Neuroscience*, *19*, 687–700.

- Leng, F., & Edison, P. (2021). Neuroinflammation and microglial activation in Alzheimer disease: Where do we go from here? *Nature Reviews. Neurology*, *17*, 157–172.
- Long, J. M., & Holtzman, D. M. (2019). Alzheimer disease: An update on pathobiology and treatment strategies. *Cell*, *179*, 312–339.
- Matschke, J., Lutgehetmann, M., Hagel, C., Sperhake, J. P., Schroder, A. S., Edler, C., Mushumba, H., Fitzek, A., Allweiss, L., Dandri, M., Dottermusch, M., Heinemann, A., Pfefferle, S., Schwabenland, M., Sumner Magruder, D., Bonn, S., Prinz, M., Gerloff, C., Püschel, K., … Glatzel, M. (2020). Neuropathology of patients with COVID-19 in Germany: A post-mortem case series. *Lancet Neurology*, *19*, 919–929.
- Murphy, C. (2019). Olfactory and other sensory impairments in Alzheimer disease. *Nature Reviews. Neurology*, *15*, 11–24.
- Olivera, E., Saez, A., Carniglia, L., Caruso, C., Lasaga, M., & Durand, D. (2023). Alzheimer's disease risk after COVID-19: A view from the perspective of the infectious hypothesis of neurodegeneration. *Neural Regeneration Research*, *18*, 1404–1410.
- Ossenkoppele, R., van der Kant, R., & Hansson, O. (2022). Tau biomarkers in Alzheimer's disease: Towards implementation in clinical practice and trials. *Lancet Neurology*, *21*, 726–734.
- Ramani, A., Muller, L., Ostermann, P.N., Gabriel, E., Abida-Islam, P., Muller-Schiffmann, A., Mariappan, A., Goureau, O., Gruell, H., Walker, A., Andrée, M., Hauka, S., Houwaart, T., Dilthey, A., Wohlgemuth, K., Omran, H., Klein, F., Wieczorek, D., Adams, O., … Gopalakrishnan, J. (2020). SARS-CoV-2 targets neurons of 3D human brain organoids. *The EMBO Journal*, *39*, e106230.
- Schwabenland, M., Salie, H., Tanevski, J., Killmer, S., Lago, M. S., Schlaak, A. E., Mayer, L., Matschke, J., Puschel, K., Fitzek, A., Ondruschka, B., Mei, H. E., Boettler, T., Neumann-Haefelin, C., Hofmann, M., Breithaupt, A., Genc, N., Stadelmann, C., Saez-Rodriguez, J., … Bengsch, B. (2021). Deep spatial profiling of human COVID-19 brains reveals neuroinflammation with distinct microanatomical microglia-T-cell interactions. *Immunity*, *54*, 1594–1610.
- Stein, S. R., Ramelli, S. C., Grazioli, A., Chung, J. Y., Singh, M., Yinda, C. K., Winkler, C. W., Sun, J., Dickey, J. M., Ylaya, K., Ko, S. H., Platt, A. P., Burbelo, P. D., Quezado, M., Pittaluga, S., Purcell, M., Munster, V. J., Belinky, F., Ramos-Benitez, M. J., … Chertow, D. S. (2022). SARS-CoV-2 infection and persistence in the human body and brain at autopsy. *Nature*, *612*, 758–763.
- Sun, B., Tang, N., Peluso, M. J., Iyer, N. S., Torres, L., Donatelli, J. L., Munter, S. E., Nixon, C. C., Rutishauser, R. L., Rodriguez-Barraquer, I., Greenhouse, B., Kelly, J. D., Martin, J. N., Deeks, S. G., Henrich, T. J., & Pulliam, L. (2021). Characterization and biomarker analyses

of post-COVID-19 complications and neurological manifestations. *Cells*, *10*, 10.

- Thaweethai, T., Jolley, S. E., Karlson, E. W., Levitan, E. B., Levy, B., McComsey, G. A., McCorkell, L., Nadkarni, G. N., Parthasarathy, S., Singh, U., Walker, T. A., Selvaggi, C. A., Shinnick, D. J., Schulte, C. C. M., Atchley-Challenner, R., Alba, G. A., Alicic, R., Altman, N., Anglin, K., … RECOVER Consortium (2023). (2023). Development of a definition of Postacute sequelae of SARS-CoV-2 infection. *Journal of the American Medical Association*, *329*, 1934–1946.
- Tsukahara, T., Brann, D. H., & Datta, S. R. (2023). Mechanisms of SARS-CoV-2-associated anosmia. *Physiological Reviews*, *103*, 2759–2766.
- van der Kant, R., Goldstein, L., & Ossenkoppele, R. (2020). Amyloidbeta-independent regulators of tau pathology in Alzheimer disease. *Nature Reviews. Neuroscience*, *21*, 21–35.
- Woo, M. S., Malsy, J., Pottgen, J., Seddiq, Z. S., Ufer, F., Hadjilaou, A., Schmiedel, S., Addo, M. M., Gerloff, C., Heesen, C., Schulze Zur Wiesch, J., & Friese, M. A. (2020). Frequent neurocognitive deficits after recovery from mild COVID-19. *Brain Commun*, *2*, fcaa205.
- Woodburn, S. C., Bollinger, J. L., & Wohleb, E. S. (2021). The semantics of microglia activation: Neuroinflammation, homeostasis, and stress. *Journal of Neuroinflammation*, *18*, 258.
- Zhang, P. P., He, Z. C., Yao, X. H., Tang, R., Ma, J., Luo, T., Zhu, C., Li, T. R., Liu, X., Zhang, D., Zhang, S., Ping, Y. F., Leng, L., & Bian, X. W. (2023). COVID-19-associated monocytic encephalitis (CAME): Histological and proteomic evidence from autopsy. *Signal Transduction and Targeted Therapy*, *8*, 24.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Qi, X., Yuan, S., Ding, J., Sun, W., Shi, Y., Xing, Y., Liu, Z., Yao, Y., Fu, S., Sun, B., Qi, X., Xia, B., Liu, F., Yi, M., Mao, J., Wan, Y., & Zheng, J. (2024). Emerging signs of Alzheimer-like tau hyperphosphorylation and neuroinflammation in the brain post recovery from COVID-19. *Aging Cell*, *00*, e14352.<https://doi.org/10.1111/acel.14352>