#### Virological characteristics of the SARS-CoV-2 JN.1 variant 1 2 Yu Kaku<sup>1#</sup>, Kaho Okumura<sup>1,2#</sup>, Miguel Padilla-Blanco<sup>3,4</sup>, Yusuke Kosugi<sup>1,5</sup>, Keiya 3 Uriu<sup>1,5</sup>, Alfredo A Hinay Jr.<sup>1</sup>, Luo Chen<sup>1,6</sup>, Arnon Plianchaisuk<sup>1</sup>, Kouji Kobiyama<sup>7,8</sup>, 4 Ken J Ishii<sup>7,8</sup>, The Genotype to Phenotype Japan (G2P-Japan) Consortium, Jiri 5 Zahradnik<sup>3</sup>, Jumpei Ito<sup>1,9</sup>, Kei Sato<sup>1,5,6,8,9,10,11</sup>\* 6 7 <sup>1</sup> Division of Systems Virology, Department of Microbiology and Immunology, 8 The Institute of Medical Science, The University of Tokyo, Tokyo, Japan 9 <sup>2</sup> Faculty of Liberal Arts, Sophia University, Tokyo, Japan 10 <sup>3</sup> First Medical Faculty at Biocev, Charles University, Vestec-Prague, Czechia 11 <sup>4</sup> Departamento de Farmacia, Facultad de Ciencias de la Salud, Universidad 12 Cardenal Herrera-CEU (UCH-CEU), CEU Universities, Valencia, Spain 13 <sup>5</sup> Graduate School of Medicine, The University of Tokyo, Tokyo, Japan 14 <sup>6</sup> Graduate School of Frontier Sciences, The University of Tokyo, Kashiwa, 15 16 Japan <sup>7</sup> Division of Vaccine Science, Department of Microbiology and Immunology, The 17Institute of Medical Science, The University of Tokyo, Tokyo, Japan. 18 <sup>8</sup> International Vaccine Design Center, The Institute of Medical Science, The 19 University of Tokyo, Tokyo, Japan 20 <sup>9</sup> International Research Center for Infectious Diseases. The Institute of Medical 21 Science, The University of Tokyo, Tokyo, Japan 22 <sup>10</sup> Collaboration Unit for Infection, Joint Research Center for Human Retrovirus 23 infection, Kumamoto University, Kumamoto, Japan 24 <sup>11</sup> CREST, Japan Science and Technology Agency, Kawaguchi, Japan 25 <sup>#</sup> Contributed equally to this study. 26 \*Correspondence: KeiSato@g.ecc.u-tokyo.ac.jp (Kei Sato) 2728

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# 30 Abstract

31 The SARS-CoV-2 BA.2.86 lineage, first identified in August 2023, is 32 phylogenetically distinct from the currently circulating SARS-CoV-2 Omicron XBB lineages, including EG.5.1 and HK.3. Comparing to XBB and BA.2, BA.2.86 33 carries more than 30 mutations in the spike (S) protein, indicating a high 34 potential for immune evasion. BA.2.86 has evolved and its descendant, JN.1 35 (BA.2.86.1.1), emerged in late 2023. JN.1 harbors S:L455S and three mutations 36 in non-S proteins. S:L455S is a hallmark mutation of JN.1: we have recently 37 shown that HK.3 and other "FLip" variants carry S:L455F, which contributes to 38 increased transmissibility and immune escape ability compared to the parental 39 EG.5.1 variant. Here, we investigated the virological properties of JN.1. 40

#### 41 **Text**

42 The SARS-CoV-2 BA.2.86 lineage, first identified in August 2023, is phylogenetically distinct from the currently circulating SARS-CoV-2 Omicron 43 XBB lineages, including EG.5.1 and HK.3. Comparing to XBB and BA.2, BA.2.86 44 carries more than 30 mutations in the spike (S) protein, indicating a high 45 potential for immune evasion.<sup>1-4</sup> BA.2.86 has evolved and its descendant, JN.1 46 (BA.2.86.1.1), emerged in late 2023. JN.1 harbors S:L455S and three mutations 47 in non-S proteins (Figure 1A). S:L455S is a hallmark mutation of JN.1: we have 48 recently shown that HK.3 and other "FLip" variants carry S:L455F, which 49 contributes to increased transmissibility and immune escape ability compared to 50 the parental EG.5.1 variant.<sup>5</sup> Here, we investigated the virological properties of 51JN.1. We estimated the relative effective reproductive number (Re) of JN.1 using 52 genomic surveillance data from France, the United Kingdom and Spain, where 53 >25 sequences of JN.1 have been reported, using a Bayesian multinomial 54 logistic model (Figures 1B, 1C, Table S3).<sup>6</sup> The R<sub>e</sub> of JN.1 in these three 55 countries was higher than that of BA.2.86.1 and HK.3, one of the XBB lineages 56 with the highest growth advantage at the end of November 2023 (Figure 1B).<sup>5</sup> 57 These results suggest that JN.1 may soon become the dominant lineage 58 worldwide. Indeed, by the end of November 2023, JN.1 has already overtaken 59 60 HK.3 in France and Spain (Figure 1C).

The *in vitro* ACE2 binding assay' showed that the dissociation constant 61 62  $(K_D)$  value of the JN.1 receptor-binding domain (RBD) is significantly higher than that of the BA.2.86 RBD (Figure 1D), suggesting that S:L455S decreases the 63 64 binding affinity to the human ACE2 receptor. In contrast, the pseudovirus assay showed that the infectivity of JN.1 is significantly higher than that of BA.2.86 65 (Figure 1E). This discrepancy (Figures 1D, 1E) would be due to the difference 66 between monomeric RBD and trimerized whole S protein (see also 67 Supplementary Discussion). We then performed a neutralization assay using 68 69 rodent sera infected with BA.2.86 or immunized with BA.2.86 S protein. In both 70 cases, the 50% neutralization titer (NT<sub>50</sub>) against JN.1 was comparable to that against BA.2.86 (Figures 1F, 1G), suggesting that S:L455S does not affect the 7172 antigenicity of BA.2.86. On the other hand, the  $NT_{50}$  of breakthrough infection (BTI) sera with XBB.1.5 and EG.5.1 against JN.1 was significantly lower than 73 that of HK.3 (2.6- to 3.1-fold) and BA.2.86 (3.8-fold) (Figures 1H, 1I). 74 Furthermore, JN.1 shows robust resistance to monovalent XBB.1.5 vaccine sera 75 compared to BA.2.86 (Figure 1J). Taken together, these results suggest that 76 JN.1 is one of the most immune-evading variants to date. Our results suggest 77 that S:L455S contributes to increased immune evasion, which partly explains the 78 79 increased R<sub>e</sub> of JN.1.

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## **Declaration of interest**

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# **Figure 1. Virological features of JN.1**

(A) Frequency of mutations in JN.1 and other lineages of interest. Only
 mutations with a frequency >0.5 in at least one but not all the representative
 lineages are shown.

(B) Estimated relative R<sub>e</sub> of the variants of interest in France, United Kingdom,
 and Spain. The relative Re of EG.5.1 is set to 1 (horizontal dashed line). Violin,
 posterior distribution; dot, posterior mean; line, 95% Bayesian confidence
 interval.

(C) Estimated epidemic dynamics of the variants of interest in France, United
 Kingdom, and Spain from April 1, 2023 to November 16, 2023. Countries are
 ordered according to the number of detected sequences of JN.1 from high to low.
 Line, posterior mean, ribbon, 95% Bayesian confidence interval.

(D) Yeast surface display affinity between the RBD of the BA.2.86 SARS-CoV-2
 variant or BA.2.86 that contained the L455S mutation and mACE2 was
 measured by yeast surface display. The dissociation constant (K<sub>D</sub>) value
 indicates the binding affinity of the RBD of the SARS-CoV-2 S protein to soluble
 ACE2 when expressed on yeast. Statistically significant differences versus
 BA.2.86 is determined by two-sided Student's *t* tests.

135 (E) Lentivirus-based pseudovirus assay. HOS-ACE2/TMPRSS2 cells were infected with pseudoviruses bearing each S protein of B.1.1 or BA.2 sublineages. 136 The amount of input virus was normalized to the amount of HIV-1 p24 capsid 137 protein. The percentage infectivity of B.1.1, BA.2 and JN.1 are compared to that 138 of BA.2.86. The horizontal dash line indicates the mean value of the percentage 139infectivity of BA.2.86. Assays were performed in guadruplicate, and a 140 representative result of four independent assays is shown. The presented data 141 are expressed as the average ± SD. Each dot indicates the result of an individual 142 replicate. Statistically significant differences versus BA.2.86 is determined by 143 two-sided Student's t tests. 144

(F-J) Neutralization assay. Assays were performed with pseudoviruses 145harboring the S proteins of B.1.1, BA.2, BA.2.86, JN.1 and HK.3. The following 146 sera were used: sera from six hamsters infected with BA.2.86 (F); sera from ten 147 mice immunized with SARS-CoV-2 BA.2.86 S (G); convalescent sera from fully 148 vaccinated individuals who had been infected with XBB.1.5 (eight 3-dose 149 150 vaccinated donors, six 4-dose vaccinated donors, four 5-dose vaccinated donors and one 6-dose vaccinated donor. 19 donors in total) (H); and EG.5.1 (one 1512-dose vaccinated donor, four 3-dose vaccinated donors, five 4-dose vaccinated 152donors, four 5-dose vaccinated donors and four 6-dose vaccinated donors. 18 153154 donors in total) (I). Assays were also performed with pseudoviruses harboring the S proteins of BA.2.86 and JN.1. The following two sera were used: 155 156vaccinated sera from fully vaccinated individuals who had not been infected (8) donors) and vaccinated sera from fully vaccinated individuals who had been 157 infected with XBB subvariants (after June, 2023) (10 donors). Sera were 158 collected before vaccination ('Pre') and 3-4 weeks after XBB.1.5 monovalent 159 vaccination ('Post') (J). Assays for each serum sample were performed in 160 triplicate to determine the 50% neutralization titer ( $NT_{50}$ ). 161

Each dot represents one NT<sub>50</sub> value, and the geometric mean and 95% 162 confidence interval are shown. The number in parenthesis indicates the 163geometric mean of NT<sub>50</sub> values. The horizontal dash line indicates the detection 164 limit (40-fold) and the number of samples with neutralization titer under the limit 165 are shown below the dash line. In F-J, statistically significant differences versus 166JN.1 were determined by two-sided Wilcoxon signed-rank tests, and p values 167are indicated in parentheses. The fold changes of NT<sub>50</sub> from that of JN.1 are 168 indicated with "X". In F and G, \*, p<0.05; \*\*, p<0.01 versus JN.1. 169

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